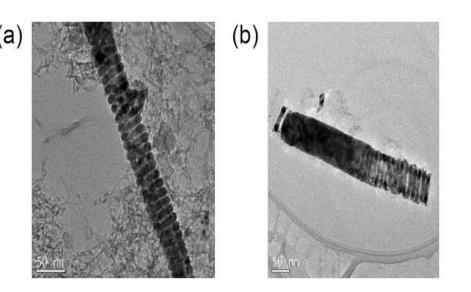
NSF NRI (1637535): Design of nanorobotics based on FePd alloy nanohelices for diagnosis and treatment of cancer



FePd nanorobot (a) nanohelix only, (b) cylinder head and helical tail.

Advantages: (i) flexible and price control under magnetic field

- (ii) Truly nano-size (10-100nm)
- (iii) MRI signal enhancement
- (iv) biocompatible
- (v) Tool for applyin mechanical stress loading to cancer cells



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Minoru Taya (PI)¹, Yasuo Kuga (Co-PI)², Donghoon Lee (Co-PI)³, Satoshi Yamamoto¹ and Satomi Takao¹

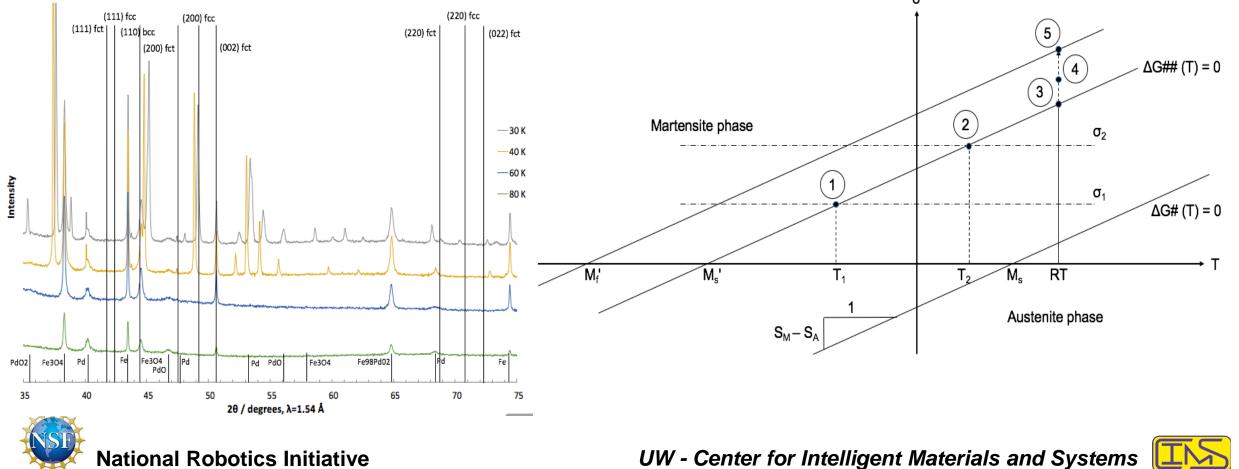
¹ Mechanical Engineering
² Electrical Engineering
³ Radiology

October 29, 2018

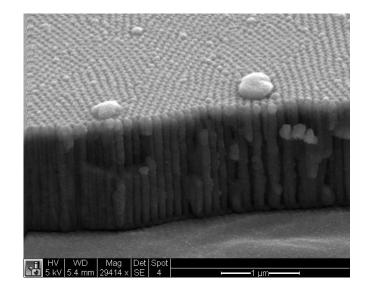


Phase Transformation Diagram of ferromagnetic shape memory alloy FePd nanomaterials,

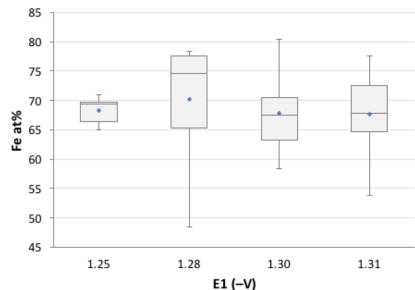
Austenite (A) to martensite(M) phase change of FePd nanohelix under decreasing temperature and increasing stress But the A-M phase boundary line of nano-sized FePd is shifted to lower temperature, as compared with bulk sized FePd, Whlie use of FePd nanorobots needs to be room temperature (RT), thus, pre-existing residual stress is needed to the A-M boundary line moving upward, shown in vertical line (see Right Figure). XRD data of synchroton (Left Figure) shows the A-Change taking place, good news !!!



Processing of FePd nanohelices is challenging 5.Back Al removal 3. Gold sputtering/evaporation on top 4. Bonding to support(ITO) by Ag Epoxy 1.PAA template with proper hole diameter 2.Silica template insertation Barrier layer Back Al 8.FePd nano-wire release 7.Electrochemical deposition



FePd composition from electrodeposition voltage





Need higher yield processing

coplymerization for ideal

Correct stoichiometry of

Fe₇₀Pd₃₀ for electroplating

nanorhelical holes in

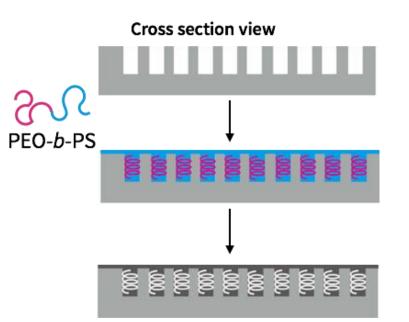
Optimized block

template

1.

2.

3.



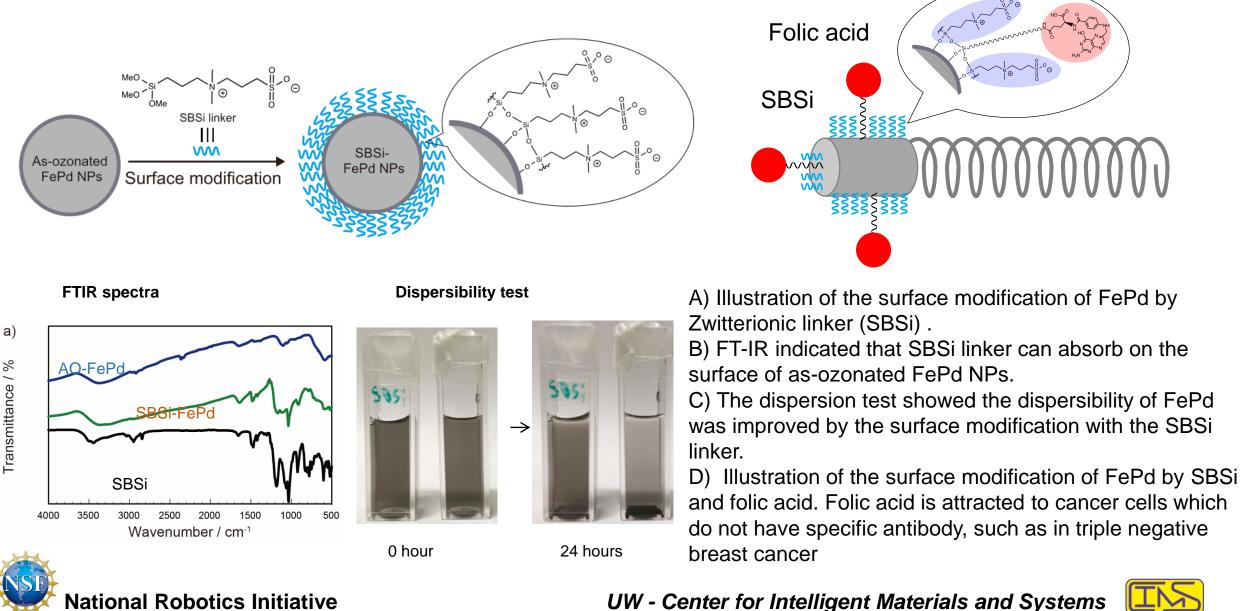
6.Barrier layer removal & hole widening

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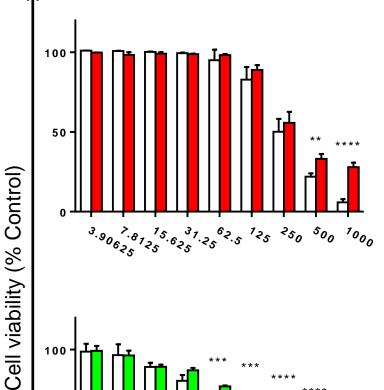
3

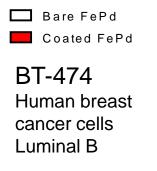
Surface coating for biocompatility and better dispersibility of FePd NRs.

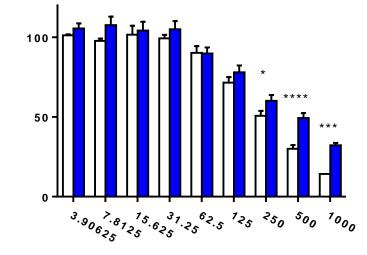




Biocompatibility assay of FePd NPs

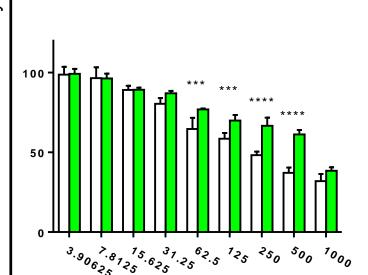






Coated FePd MDA-MB-231 Human breast cancer cells Triple negative

Bare FePd



| Bare FePd |
|-------------|
| Coated FePd |

M6 Murine breast cancer cells Triple negative

Concentration of bare or coated Fe₇₀Pd₃₀ NPs (µg/ml)

Three cell lines were cultured with bare or SBSi-coated $Fe_{70}Pd_{30}$ NPs for 48 hr and cell viability was measured by XTT assay.

| Table 1. GI50 of bare and coated $Fe_{70}Pd_{30}$ NPs (µg/ml) | | | | | | | | |
|---|------|--------|--|--|--|--|--|--|
| Cell line | Bare | Coated | | | | | | |
| BT-474 | 252 | 218 | | | | | | |
| MDA-MB-231 | 217 | 223 | | | | | | |
| M6 | 226 | 748 | | | | | | |

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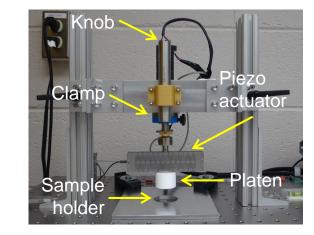
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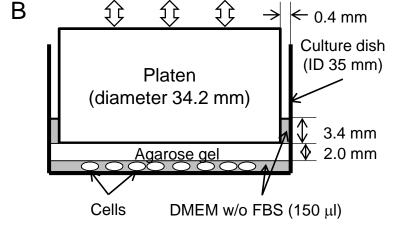
5

Mechanical Stress-induced Cell Death (MSICD)

Mechanical stress was applied to cancer cells macroscopically and we assessed cell damages.



А



Equation: $\delta(t) = \alpha/2^*(\sin(2^*pi^*f)+1)^*(1-e^{-t/\tau 1}) + \beta^*(1-e^{-t/\tau 2}))$

Setting parameters of mechanical stress loading testing

- Agarose gel: 3.0 %, ~2 mm thickness
- f: 30 Hz (the displacement frequency)
- α : 40-130 μ m (the displacement amplitude)
- t: 210-450 sec (duration)

- τ 1: 50 sec (the time constant for the ramp-up stage)
- $\tau 2$: 50 sec (the time constant for the additional compressive load) •

С

350

200

້ອງ 150

Displace 50

50

35

30

25

Ê 20

15 Force

10

100

100

200

200

Time (sec)

300

300

(un 300 250

 β : 200, 220 μ m (the additional compressive displacement to prevent tensile loading) ٠

Initial platen movement was operated manually to reach the surface of gels (~0.2 N), then started the program in LabVIEW. DMEM, Dulbecco's modified Eagle's medium; FBS, fetal bovine serum.

Cells used in the experiments are breast cancer cell lines, BT-474 (luminal B) and MDA-MB-231 (triple negative).

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6

→ u max

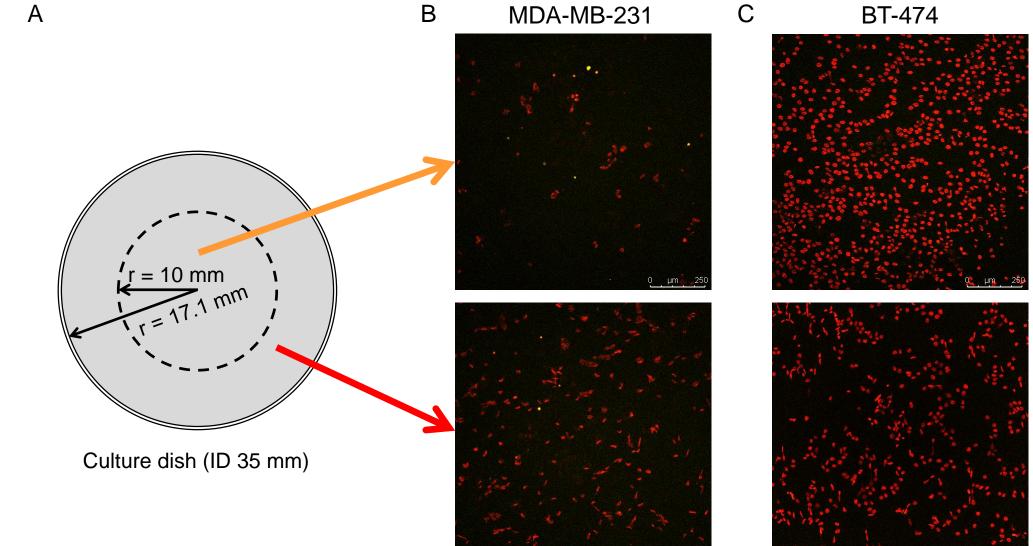
F max

F min

Amplitude of

force, ΔF

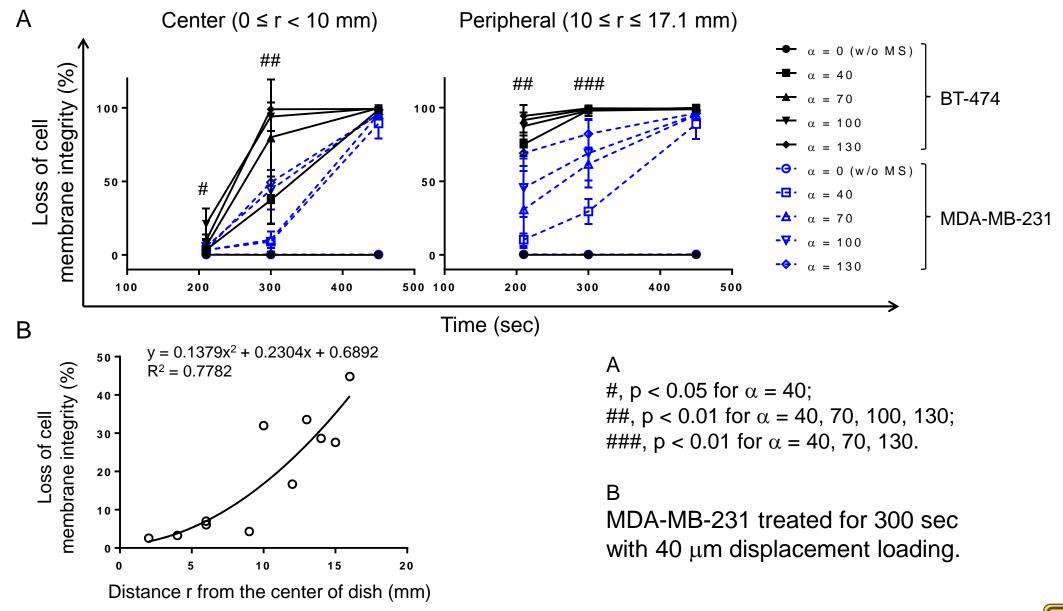
MSICD: two regions in culture dish







Necrosis-based MSICD



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8

MRI relaxation time (T2= 1/R2) of FePd nanoparticles, which are found to enhance MRI signals

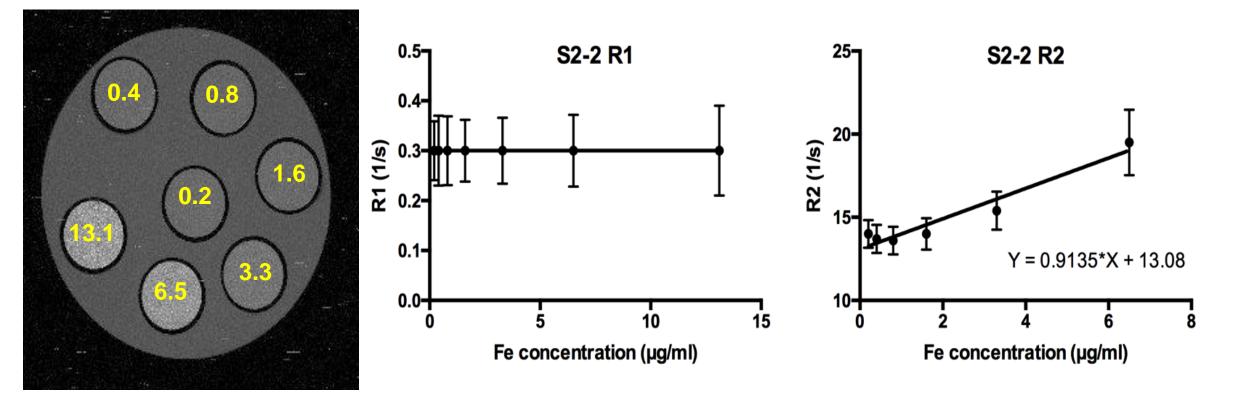


Figure #. T2 weighted images containing different iron concentrations in μ g/ml (left) and R1 (middle) and R2 (right) as a function of iron concentration.

The slope of R2- concentration graph is higher than that of R1, thus, use of R2 data is better suited for our FePd nanorobots to detect their location inside mice or human.





Nanorobot Swimming Speed and Swim Time Model

 $F = VM * \nabla H_{\chi}$ T = VM * H

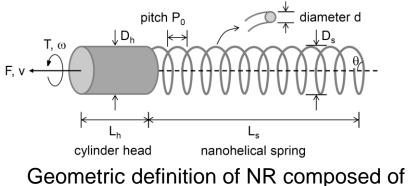
Where, $H \cong H_y \cong H_z$ (*Rotational EM Field*) and relationship (34)

 $\begin{bmatrix} V \\ T \end{bmatrix} = \begin{bmatrix} \alpha & \beta \\ -\beta & \gamma \end{bmatrix} \begin{bmatrix} F \\ \omega \end{bmatrix}$

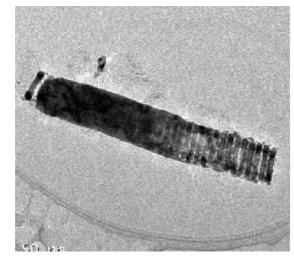
With respective element computed by definition result in:

$$\binom{\mathbf{v}}{T} = \begin{bmatrix} 1.76 \times 10^7 & \left[\frac{m}{N.s}\right] & 1.391 \times 10^{-9}[m] \\ -1.391 \times 10^{-9}[m] & 3.378 \times 10^{-23}[N.s.m] \end{bmatrix} \binom{F}{\omega}$$

With F and T computed we can obtain $\omega_{step out}$ as 51,125 rad/s. Thus exceeding the step out angular velocity is impossible for our driver system to achieve due to high induction and overheating from flux leakage.



head and nanohelix tail.



TEM photo of a FePd nanorobot made of cylinder head and nanohelix tail.



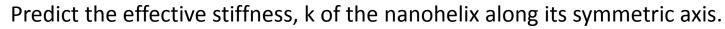
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Nanorobot Swimming Model: velocity and spring constant

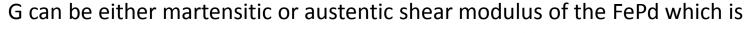
| Case | $D_h(nm)$ | $L_h(nm)$ | D _s (nm) | d = 2r (nm) | L _s (nm) | n | Velocity (mm/s) | Swim Time (Hours) | k _{mart} (N/m) | k _{aust} (N/m) |
|------|-----------|-----------|---------------------|--------------------|---------------------|----|--------------------|----------------------|-----------------------------------|----------------------------|
| 0 | 60 | 200 | 60 | 10 | 250 | 16 | 0.000525 | 12.7 | 0.00392 | 0.01305 |
| 1 | 60 | 200 | 200 | 20 | 1000 | 19 | 0.00161 | 3.46 | 0.00141 | 0.00470 |
| 2 | 60 | 200 | 1000 | 50 | 20000 | 76 | 0.0086 | 0.65 | 0.00011 | 0.000367 |
| 3 | 200 | 600 | 200 | 20 | 1000 | 19 | 0.00157 | 3.53 | 0.00141 | 0.00470 |
| 4 | 1000 | 3000 | 1000 | 50 | 20000 | 76 | 0.00854 | 0.65 | 0.00011 | 0.000367 |
| 5 | 1000 | 3000 | 2000 | 50 | 10000 | 19 | 0.018 | 0.32 | 0.000055 | 0.000184 |
| 6 | 1000 | 3000 | 1000 | 50 | 10000 | 38 | 0.0085 | 0.65 | 0.00022 | 0.000734 |

Comparison of swimming velocity of NRs and Swim time of NRs of different cases are done based on similar EM driving condition assumed for all cases which are:

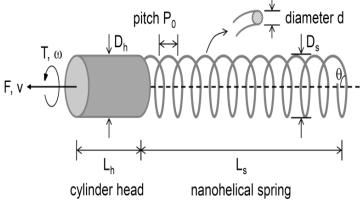
- f = 50Hz rotational EM field, l = 2cm swimming length,
- 14mT rotational maximum magnetic field density and
- 0.2 *T*/*m* magnetic field density gradient.



 $k = \frac{d^4 G}{8D_s^3 n}$



dependent on its shape memory effect.

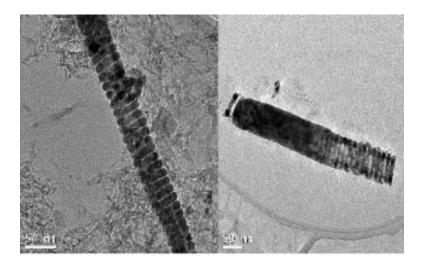




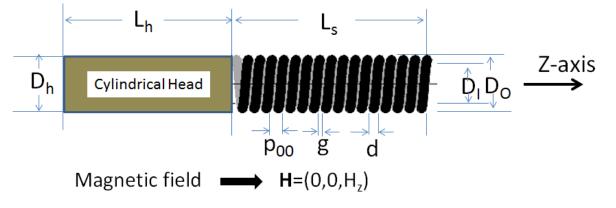
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Molecular Dynamics Model Simulation for Nano-motions of FePd nanohelix robots under applied magnetic **field**



Left : nanohelix only Right: nanorod(head) connected to nanohelix(tail)



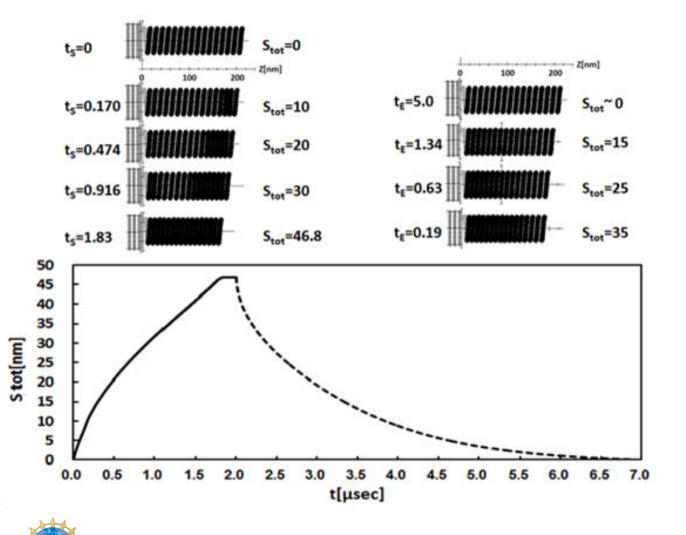
Molecular dynamics model is developed to simulate the nanomotions of FePd nanorobots under applied magnetic field, for two cases, (i) on-aixs magnetic field, and (ii) off-axis magnetic field

Taya et al (2017), J. Appl. Phys., **121**, 154302





Molecular dynamics (MD) model Simulations for the robot made of nanorod and nanohelix with left end fixed under on-axis magnetic field



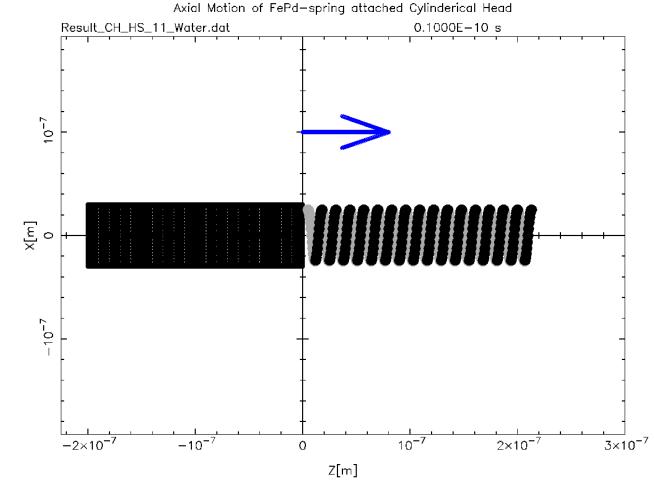
Fast nano-motions of FePd NRs under applied magnetic field on(solid curve in left graph) and off (dashed curve)

Taya et al (2017), J. Appl. Phys., **121**, 154302





Video: Molecular Dynamics model simulation of FePd NR under on-axis magnetic field

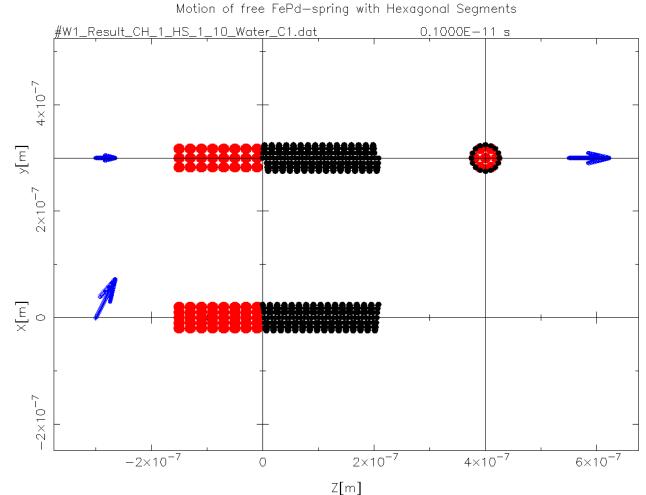




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Video: Molecular dynamics model to simulate the nanomotions of FePd nanorobots under off-axis applied magnetic field

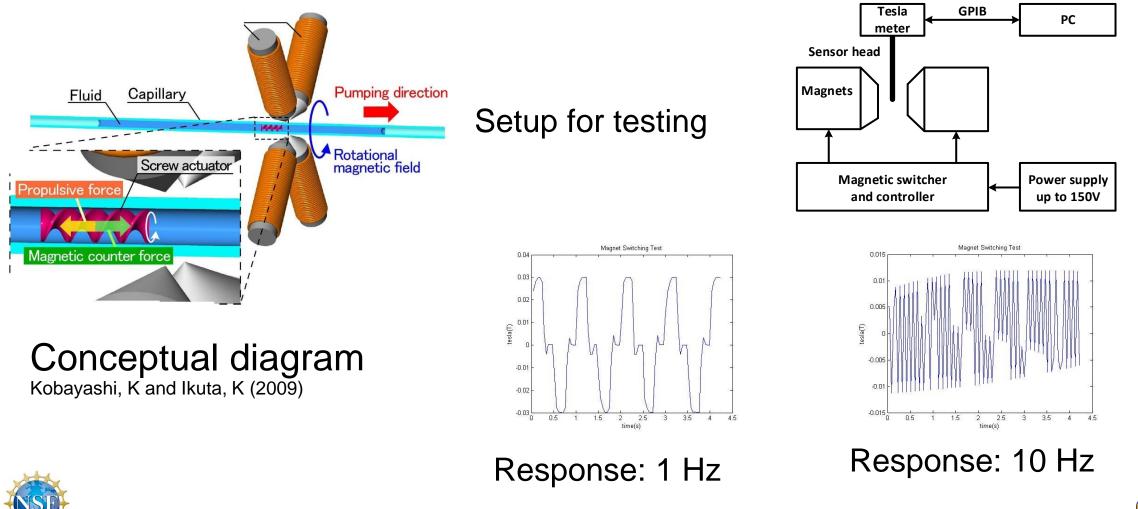






2D Helmholtz coil system for swimming of FePd nanorobots

Use of lower frequency is better for more accurate control and also adequate for inducing cancer cell death





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Acknowledgements:

UW collaborators:

Profs. Y. Kuga(EE), D. Lee(Radiology), S.Stanton(Medicine), R. Schick (Physics)

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